

# EFFECT OF JUVENILE HORMONE ANALOGOUS ON COTTON LEAFWORM

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## Abstract

The effects of the juvenile hormone analogues pyriproxyfen [2-(1-methyl 1-2-(phenoxyphenoxy) ethoxy) pyridine] on total protein, amino acid transferases, acid and alkaline phosphatases of *Spodoptera littoralis* were studied after feeding the 4<sup>th</sup> and 6<sup>th</sup> instar larvae on the LC<sub>50</sub> (225 ppm) of the tested compounds. The results showed that pyriproxyfen caused a significant reduction in the level of the total protein at all time intervals tested (48, 72, 96 and 144 hours). The data revealed that there was a significant increase in activity of glutamic Oxaloacetic transaminase (GOT) during the different time intervals comparing with the untreated check. While there was a significant reduction in the level of glutamic pyruvic transaminase (GPT) after 48 and 72 hours in both 4<sup>th</sup> and 6<sup>th</sup> instar larvae. In contrast, at 96 and 144 hours, there was a significant increase. Pyriproxyfen also cause a significant decrease in activity of alkaline phosphatase (alkapase) and acid phosphatase at different time intervals.

## Introduction

The juvenile hormone analogues as insect growth regulators have gained attention for insect control, because of their unique mode of action, that differ from the conventional insecticides. Recently, several compounds of this group have been developed which are still much more active such as pyriproxyfen. In addition to their effect on the reproductive system, these compounds disrupt the insect development from egg, through instar stages, to pupa and imago by controlling the timing for molting of the insect exoskeleton (Abdallah, et. al., 1974 and 1975). On the other hand, understanding of resistance mechanism of insects toward insecticides is an important factor in the IPM programmes. It is necessary to determine the vital role of the enzyme system of insect in destroying such compounds. This phenomena have been studied by (Abdel-Fattah et. al., 1986; Abdel-Hafez et. al., 1988; Gadallah et. al., 1990 and 1994.

The purpose of this work was carried out to study the effects of the juvenile hormone mimic pyriproxyfen on the activities of some biochemical aspects of *Spodoptera littoralis* that main problem of cotton and other crops in Egypt.

## Materials and Methods

### Test Insect

Laboratory strain of the cotton leafworm, *Spodoptera littoralis* (Boisd.) which has been reared in the laboratory for ten generation away from any insecticide contamination was used in these studies. The larval instars were fed on castor bean leaves, *Ricinus communis* (L.) After pupation, the pupae were placed in a wide glass jars until the adults emergence. Then, the emerged adults were supplied with a piece of cotton wetted with 10% sugar solution and branches of tafla (*Nerium oleander*) as suitable site for oviposition (El-Defrawi et. al., 1964). All stages were reared and treated under constant conditions of 25±2 °C and 65± R.H.

### Chemicals Treatment Procedure

The 4<sup>th</sup> and 6<sup>th</sup> instar larvae were fed on castor oil leaves treated with different concentrations of the tested compound during the preliminary experiments. Then LC<sub>50</sub> values were determined from the mortality regression lines (Finney, 1952).

### Collection of Haemolymph Samples

Haemolymph samples were collected from both 4<sup>th</sup> and 6<sup>th</sup> instar larvae which fed on castor oil leaves previously treated with LC<sub>50</sub> (225 ppm) of the tested compound. The haemolymph samples were collected at 48, 72, 96 and 144 hour after transferring to untreated leaves. The procedure described by Chen and Levenbook, 1960 was followed. Collected haemolymph was kept inconvenient small well stoppered glass (7 x 0.5 cm). After being centrifuged at 2500 r.p.m. for 10 minutes, the samples were stored in refrigerator at - 20°C.

### Protein Determination

Total protein in the haemolymph samples was determined according to Lowry et. al. (1951).

### Determination of Transaminase Activities

Determination of glutamic oxaloacetic transaminase (GOT) and glutamic pyruvic transaminase (GPT) activities were carried out according to Reitman and Frankel (1957).

### Determination of Acid and Alkaline Phosphatase

The activity of acid and alkaline phosphatase enzymes were determined according to the method of Shinowara et. al. (1942).

### Statistical Analysis

The means and standard deviation were calculated for each experiment and the data were compared using the ANOVA test according to Snedecor (1971).

## Results and Discussion

Feeding the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *Spodoptera littoralis* on castor oil leaves previously treated with LC<sub>50</sub> of pyriproxyfen caused a significant decrease in the level of the total protein at all time intervals tested for the 4<sup>th</sup> and 6<sup>th</sup> instar larvae comparing with the untreated check as shown in Table (1).

The decrease of total protein might reflect the decrease in the activity of various enzymes. The pyriproxyfen could be considered as inhibitory agents for protein synthesis in *S. littoralis*. These results are in agreement with Abdel-Hafez et. al. (1988) and Gadallah et. al. (1994).

Data in Table (2) indicated that pyriproxyfen treatment showed a significant increase in the level of glutamic oxaloacetic transaminase (GOT) activity after 48, 96 and 144 hours of feeding the 4<sup>th</sup> instar larvae on treated castor bean leaves with LC<sub>50</sub>, while highly significant decrease was found after 72 hours of treatment in comparison with the untreated larvae.

As for the 6<sup>th</sup> instar larvae, application with the LC<sub>50</sub> revealed significant decrease in GOT activity after 48 and 72 hours, in contrast the enzyme activity was increased after 96 and 144 hours post-treatment comparing with untreated check.

Considering alanine amino transferase activity (GPT), Table (3) showed that there was a significant reduction in the level of glutamic pyruvic transaminase (GPT) after 48 and 72 hours in both 4<sup>th</sup> and 6<sup>th</sup> instar larvae. In contrast, at 96 and 144 hours, there was a significant increase.

It is clear from our results that pyriproxyfen revealed an pronounced increase in transaminases (GOT & GPT) at most time intervals after treatment. Since transferases are group of enzyme which responsible for maintenance of the balance amino acid pool in insects. Consequently, the degradation in hemolymph protein increased resulted in reduction in protein content. It has been reported by many workers that the level of amino acid transferases varies with the amount of protein synthesis (Chen, 1966; Gilbert, 1967; Gadallah et. al., 1994). Zidan et. al. (1996) found that

immediately after treatment, *B. thuringiensis* had the most significant effect on reduction of protein content of the larvae, but KZ oil showed the highest overall reduction 72 h after treatment. Pyriproxyfen resulted in the best immediate inhibitory effect on AchE [acetylcholinesterase] whereas *B. thuringiensis* and KZ oil caused better latent effects. Pyriproxyfen and *B. thuringiensis* caused considerable reduction in ACP (acid phosphatase) activity (72 h) after treatment. The inhibition of GPT [alanine aminotransferase] was greatest within 24 h for all 3 insecticides used.

The effect of pyriproxyfen on the activities of haemolymph acid phosphatase (Acpase) and alkaline phosphatase (Alkpase) in 4<sup>th</sup> and 6<sup>th</sup> instar larvae were determined at different time intervals, as shown in Tables (4 and 5).

The data revealed that pyriproxyfen caused a significant reduction in the activities of Acpase enzyme at all time intervals treated for the two instar larvae. Alkpase activities showed a different trend. There was a significant difference at all time interval except at 24 hours after treatment.

The present results agree with the finding of Abdel-Hafez et. al. (1988) and El-Kordy et. al. (1995).

### References

Abdallah, M.D., Zaazou, M.H. and El-Tantawy, M.A. 1974. The morphogentic activity of a juvenile hormone analous in *Spodoptera littoralis* (Boisd.) *Toxicology.*, 2:339-347.

Abdallah, M.D., Zaazou, M.H. and El-Tantawy, M.A. 1975. Reduction in fecundity of adult females and hatchability of egg larvae of *Spodoptera littoralis* (Boisd.) after exposure of pupae and eggs to juvenile hormone and analogous. *Z. Angew. Ent.*, 78:176-181.

Abdel-Fattah, M.S., El-Mallah, M. A. and Shaaban, M.N. 1986. Effect of diflubenzuron and triflumeron on the activity of esterases in susceptible and profenofos-resistant strains of *Spodoptera littoralis* (Boisd.). *Bull. Ent. Soc. Egypt. Econ. Ser.*, 15:221-227.

Abdel-Hafez, M.M., M.N. Shaaban, M.A., El-Malla, M. Farag, and A.M. Abd El-Kawy. 1988. Effect of insect growth regulators on the activity of transaminase with reference to protein and amino acids in the Egyptian cotton leafworm, *Spodoptera littoralis* (Boisd.) *Minia J. Agric. Res. & Dev.*, 10:1391-1040.

Chen, P.S. and L. Levenbook. 1960. Studies on the haemolymph proteins of the blowfly *Phormia regina* L. Changes in ontogenetic patterns. *J. Insect Physiol.*, 12:1595-1609.

El-Kordy, M.W., A.I. Gadallah, M.G. Abbas and S.A. Mostafa. 1995. Changes in phosphatases and carbohydrases during the different stages of *Earias insulana* (Boisd.). *Al-Azhar J. Agric. Res.*, 22:217-228.

El-Defrawi, M.E., Topozada, A., Monsour, N. and Zeid, M. (1964): Toxicological studies on the Egyptian cotton leafworm, *Prodenia littura* (L.). I. Susceptibility of different larval instars of *Prodenia* to insecticides. *J. Econ. Entomol.*, 57:591-593.

Finney, D.J. 1952. *Probit Analysis* (second edition), Cambridge Univ. Press., 318 pp.

Gadallah, A.I., M.W. El-Kordy. M.G. Abbas and S.A. Mostafa. 1994. Effect of flufenoxuron, teflubenzuron and pyriproxyfen on some dehydrogenase of *S. littoralis*. *Al-Azhar J. Agric. Res.*, 20:347-361.

Gadallah, A.I., G.M. Moawad, M.G. Abbas and S. Emam. 1990. Biological and biochemical effects of the juvenile hormone mimic S-31193 on the American bollworm *Heliothis armigera* (Hbn.) (Lepidoptera: Noctuidae). *Bull. Ent. Soc. Egypt, Econ. Ser.*, 18:125-136.

Gilbert, L.I. 1967. Biochemical corelation in insect metamorphosis. *Comp. Biochem.*, 28:199-252.

Lowry, O.H., N.H. Bosebrough, A.L. Farr and R.S. Randall. 1951. Protein measurement with the folinphenol reagent. *J. Biol. Physiol.*, 5:129-172.

Reitman, S. and S. Frankel. 1957. Colourimetric method for aspartate and alanine transaminase. *Amer. J. Clin. Pathol.*, 28:56.

Shebl, D.E.A.F. 1979. Physiological and biochemical studies on the American bollworm. M.Sc. Thesis, Fac. Agric., Cairo Univ., Egypt.

Shinowara, G.Y., L.M. Jones and H.L. Reinhart. 1942. The estimation of serum inorganic phosphate and acid and alkaline phosphatase activity. *J. Biol. Chem.*, 142:921-927.

Snedecor, G.W. 1971. *Methods of Statistical Analysis*. Iowa State Univ. Press, Ames, Iowa, U.S.A.

Sridhara, S. and J.V. Bhat. 1963. Alkaline and acid phosphatases of the silk worm, *Bombyx mori* L. *Insect Physiol.*, 9:693-701.

Zidan-ZH; Moawad-GM; Galallah-AI; El-Sweeki-FE (1996): Biochemical aspects of the cotton leafworm larvae *Spodoptera littoralis* (Boisd.) as affected by soft nontoxic insecticides. *Proceedings: 6<sup>th</sup> Conf. Agric. Dev. Res.* 17-19 December 1996, Cairo. *Annals-of-Agricultural-Science-Cairo*. 1996, No. Special Issue, 233-244.

Table 1. Changes in protein contents in the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *S. littoralis* after treated with LC<sub>50</sub> values of pyriproxyfen with leaf dipping technique.

*h.p.t.	Protein contents (µg/g b. wt. ± S.D.)			
	48 hr	72 hr	96 hr	144 hr
<b>4<sup>th</sup> instar</b>				
Pyriproxyfen	26.5±1.5a	28.43±1.4a	29.43±1.3a	30.11±1.4a
Check	33.8±1.9b	24.28±1.8a	36.25±1.7b	50.32±2.1b
<b>6<sup>th</sup> instar</b>				
Pyriproxyfen	27.44±1.7a	36.55±1.3a	34.14±1.7a	35.24±2.1a
Check	35.22±1.5b	41.32±1.3b	45.32±1.2b	44.33±1.1b

- Figures followed by the same letters are statistically insignificant (P>0.05), those followed by the different letter are significantly different (P>0.5).

\*h.p.t.: hours post treatment.

Table 2. Changes in GOT haemolymph contents in the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *S. littoralis* after treated with LC<sub>50</sub> values of pyriproxyfen with leaf dipping technique.

*h.p.t.	GOT haemolymph contents (µg/g b. wt. ± S.D.)			
	48 hr	72 hr	96 hr	144 hr
<b>4<sup>th</sup> instar</b>				
Pyriproxyfen	26.6±0.8a	19.4±1.0a	48.3±2.3a	41.4±2.2a
Check	9.3±0.7b	50.3±2.3b	36.3±1.7b	32.5±2.1b
<b>6<sup>th</sup> instar</b>				
Pyriproxyfen	29.4±1.1a	11.9±0.8a	41.61±1.3a	42.3±1.4a
Check	31.5±1.1b	41.3±2.1b	33.2±1.3b	33.6±1.1b

- Figures followed by the same letters are statistically insignificant (P>0.05), those followed by the different letter are significantly different (P>0.5).

\*h.p.t.: hours post treatment.

Table 3. Changes in GPT haemolymph contents in the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *S. littoralis* after treated with LC<sub>50</sub> values of pyriproxyfen with leaf dipping technique.

GPT haemolymph contents ( $\mu\text{g/g b. wt.} \pm \text{S.D.}$ )				
*h.p.t.	48 hr	72 hr	96 hr	144 hr
<b>4<sup>th</sup> Instar</b>				
Pyriproxyfen	35.5 $\pm$ 1.9a	14.5 $\pm$ 1.4a	34.4 $\pm$ 1.3a	45.3 $\pm$ 1.2a
Check	115.3 $\pm$ 2.3b	46.5 $\pm$ 2.9b	28.3 $\pm$ 0.9b	32.6 $\pm$ 1.0b
<b>6<sup>th</sup> Instar</b>				
Pyriproxyfen	26.4 $\pm$ 0.8a	12.4 $\pm$ 1.2a	49.5 $\pm$ 1.1a	55.4 $\pm$ 1.4a
Check	32.5 $\pm$ 1.4b	43.1 $\pm$ 2.4b	38.3 $\pm$ 0.7b	36.3 $\pm$ 1.5b

- Figures followed by the same letters are statistically insignificant ( $P>0.05$ ), those followed by the different letter are significantly different ( $P>0.5$ ).

\*h.p.t.: hours post treatment.

Table 4. Changes in acid phosphatase contents in the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *S. littoralis* after treated with LC<sub>50</sub> values of pyriproxfen with leaf dipping technique.

Acid phosphate contents ( $\mu\text{g/g b. wt.} \pm \text{S.D.}$ )				
*h.p.t.	48 hr	72 hr	96 hr	144 hr
<b>4<sup>th</sup> instar</b>				
Pyriproxyfen	8.3 $\pm$ 0.6a	6.9 $\pm$ 0.7a	3.4 $\pm$ 0.5a	2.3 $\pm$ 0.6a
Check	14.3 $\pm$ 0.8b	11.5 $\pm$ 0.8b	11.4 $\pm$ 0.7b	12.32 $\pm$ 0.7b
<b>6<sup>th</sup> instar</b>				
Pyriproxyfen	11.4 $\pm$ 0.7a	7.3 $\pm$ 0.7a	5.8 $\pm$ 0.7a	3.3 $\pm$ 0.8a
Check	16.8 $\pm$ 0.9b	12.9 $\pm$ 1.0b	13.4 $\pm$ 0.9b	11.1 $\pm$ 0.9b

- Figures followed by the same letters are statistically insignificant ( $P>0.05$ ), those followed by the different letter are significantly different ( $P>0.5$ ).

\*h.p.t.: hours post treatment.

Table 5. Changes in alkaline phosphatase contents in the 4<sup>th</sup> and 6<sup>th</sup> instar larvae of *S. littoralis* after treated with LC<sub>50</sub> values of pyriproxyfen with leaf dipping technique.

Alkaline phosphate contents ( $\mu\text{g/g b. wt.} + \text{S.D.}$ )				
*h.p.t.	48 hr	72 hr	96 hr	144 hr
<b>4<sup>th</sup> instar</b>				
Pyriproxyfen	6.32 $\pm$ 0.4a	4.21 $\pm$ 0.3a	8.91 $\pm$ 0.6a	9.5.3 $\pm$ 0.6a
Check	8.45 $\pm$ 0.6a	8.92 $\pm$ 0.5b	12.14 $\pm$ 0.8b	15.52 $\pm$ 1.1b
<b>6<sup>th</sup> instar</b>				
Pyriproxyfen	7.92 $\pm$ 0.7a	5.2 $\pm$ 0.4a	8.4 $\pm$ 0.7a	10.1 $\pm$ 0.9a
Check	10.13 $\pm$ 1.1a	9.4 $\pm$ 0.8b	13.2 $\pm$ 0.9b	14.3 $\pm$ 1.2b

- Figures followed by the same letters are statistically insignificant ( $P>0.05$ ), those followed by the different letter are significantly different ( $P>0.5$ ).

\*h.p.t.: hours post treatment.