

LABORATORY EVALUATION OF SELECTED INSECTICIDES ON FIELD-COLLECTED POPULATIONS OF BOLLWORM AND TOBACCO BUDWORM LARVAE

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Abstract

Bollworm (CEW) and tobacco budworm (TBW) larvae and adults were collected from a variety of host crops and evaluated for susceptibility to MVP II[®], cypermethrin and spinosad (Tracer[®]) during the 2010 season. Results were compared to historical data collected throughout a sixteen-year study period beginning in 1995. As expected, CEW larvae were less susceptible to the effect(s) of MVP II[®] than were TBW larvae. Throughout the study period, the susceptibilities of CEW and TBW larvae to MVP II[®] (though variable) have decreased. The mean 2010 MVP II[®] LC₅₀ value for CEW was ca. 10-fold higher than 2009 values and the mean 2010 MVP II[®] LC₅₀ value for TBW was ca. 4.5-fold higher than values recorded during the 2009 season. Although cypermethrin remains an effective insecticide to control CEW larvae, average LC₅₀ values were highest during the 2010 season and ca. 8-fold higher than LC₅₀ values recorded in the mid-1990s. The effectiveness of cypermethrin for the control of TBW larvae has declined throughout the fifteen-year study period also. The average LC₅₀ values for cypermethrin against TBW larvae collected during the 2010 season were ca. 48-fold higher than the LC₅₀ value obtained for a pyrethroid-susceptible laboratory strain and ca. 17-fold higher than the average LC₅₀ values obtained during the mid-1990s. Decreases in the susceptibilities of CEW and TBW populations were confirmed by the use of topical application bioassays and adult vial tests. Spinosad (Tracer[®]) remained highly effective against CEW and TBW larvae throughout the study period; however, the highest average LC₅₀ values to date were obtained during the 2009 and 2010 seasons.

Introduction

The bollworm (CEW; *Helicoverpa zea*) and the tobacco budworm (TBW; *Heliothis virescens*) are two of the more economically important pests of cotton in the United States. Because CEW and TBW populations have developed resistance to many of the insecticides used for their control, it is essential that research efforts and agricultural practices be devoted to the preservation of those insecticides that are still effective and to the development of new replacement compounds and technologies. Programs to monitor insecticide susceptibilities of field-collected populations of CEW and TBW are critical to the development of those effective management strategies. Samples of CEW and TBW populations were collected from cotton, tobacco, peanut and corn fields throughout Georgia during the summer of 2010. Larvae from those field-collected samples were assayed for susceptibility to a variety of insecticides using treated-diet and topical application bioassays; adults were evaluated using an adult vial test bioassay. Results were compared to baseline data collected between 1995-1999, 2003-2005, and 2009.

Materials and Methods

The counties from which CEW and TBW have been collected throughout the study period are shown in Figure 1. During the 2010 season, 7 CEW and 5TBW populations were collected from 4 counties including Carroll, Jeff Davis, Miller and Tift. Field-collected CEW and TBW moths or larvae were transported to facilities at the University of West Georgia. Larvae were transferred to a pinto bean/wheat germ, agar-based diet, and adults were placed in mating cages to produce adequate numbers of larvae for testing. Larvae and adults were maintained at 27°C, LD 14:10 and ca. 40% RH. The insecticides used were MVP II[®] (19.1% A.I., Monsanto Corporation, St. Louis, MO); cypermethrin (94.3% A.I., FMC Corporation, Princeton, NJ); and spinosad (91.3% A.I., Dow AgroSciences, Indianapolis, IN).

Larvae were evaluated using a modified insecticide-treated diet bioassay or by topical application; adults were evaluated using an adult vial test (AVT) protocol. For the insecticide-treated diet assay, an insecticide test solution (100 µl) was added to 50 mL of liquefied pinto bean/wheat germ, agar-based diet at ca. 57°C while mixing with a variable speed stirrer. The insecticide-treated diet (ca. 2.5 mL) was distributed into 1 oz. clear plastic medicine cups. The treated diets were allowed to cool and gel. One neonate or one late 2nd instar larva (depending upon the insecticide being evaluated) was added to each cup, and mortality was monitored over a 4 day period. For the topical application bioassay, a 1 µL droplet an insecticide solution or acetone (control) was applied to the dorsal

thorax of a 4th instar larva (ca. 35 mg). Mortality was assessed after a 48 h exposure period. For the adult vial test, a single moth was placed in an insecticide-treated or acetone-treated (control) vial. Mortality was assessed after a 24h exposure period. Mortality was defined as the inability of the larva to move across the diet surface when probed or for a moth to fly a distance of 1 meter when dropped from a 2 meter height. During the treatment period, the larvae and adults were held in an environmental chamber at 27°C, LD 14:10 and ca. 40% RH.

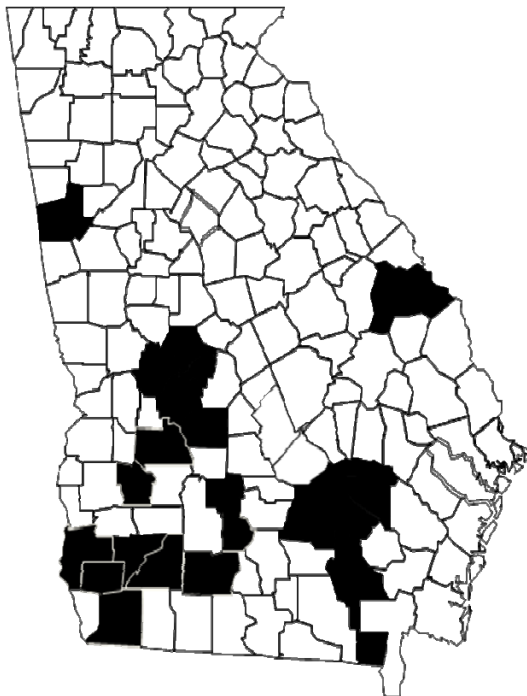


Figure 1. Bollworm and tobacco budworm collection sites.

Results and Discussion

As expected, MVP II[®] was less effective against CEW larvae as compared to TBW larvae (Tables 1-4; Figures 2, 3, 4 and 5). The average CEW LC₅₀ value were ca. 40-fold greater than the average TBW LC₅₀ value. Although isolated CEW and TBW populations exhibited high levels of survival following exposure to MVP II[®], the average CEW and TBW LC₅₀ values for the 2010 season were ca. 4-fold greater than average CEW and TBW LC₅₀ values obtained during the 2009 season. The highest CEW LC₅₀ values (> 500 ppm) were recorded during the 2010 season and were, on average, more than 5-10-fold higher than the LC₅₀ values obtained during the previous sixteen years.

Table 1. Susceptibilities of field-collected bollworm populations to MVP II[®], spinosad (Tracer[®]), and cypermethrin using treated diet, topical application, and adult vial test (AVT) bioassays—2010.

Colony	Treatment	<u>Diet</u> LC ₅₀ (C.I.; Slope), ppm	<u>Topical*</u> LC ₅₀ (C.I.; Slope), µg/g	<u>AVT**</u> LC ₅₀ (C.I.; Slope), µg/vial
TIF A 10	MVP II [®]	795.0 (376.0-2210; 0.71)		
TIF B 10		134.0 (99.4-189; 1.09)		
TIF E 10		855.0 (458.0-2315; 0.84)		
TIF F 10		536.0 (258.0-1735; 0.73)		
UWG 10		152.0 (103.0-227.0; 1.84)		
TIF A 10	Spinosad	0.32 (0.24-0.45; 2.72)		
TIF C 10		0.60 (0.36-1.03; 2.16)		
TIF F 10		0.78 (0.59-1.04; 1.76)		
TIF A 10	Cypermethrin	9.14 (7.10-11.8; 3.11)	-----	-----
TIF B 10		16.1 (12.0-22.0; 2.70)	4.71 (2.63-8.94; 2.63)	-----
TIF C 10		7.54 (5.26-10.6; 2.87)	-----	4.88 (3.74-6.43; 2.34)
TIF F 10		18.8 (13.8-25.7; 1.77)	-----	-----
UWG 10		7.15 (5.81-8.83; 2.31)	-----	4.70 (3.32-6.88; 2.59)

* larval weight: ~35 mg **Adult Vial Test (males and female adults evaluated)

Table 2. Susceptibilities of field-collected tobacco budworm populations to MVP II[®], spinosad (Tracer[®]), and cypermethrin using treated diet, topical application, and adult vial test (AVT) bioassays—2010.

Colony	Treatment	<u>Diet</u> LC ₅₀ (C.I.; Slope), ppm	<u>Topical*</u> LC ₅₀ (C.I.; Slope), µg/g	<u>AVT**</u> LC ₅₀ (C.I.; Slope), µg/vial
TIF 10	MVP II [®]	2.33 (1.63-3.22; 1.09)		
HAZ 10		4.07 (2.54-6.27; 1.07)		
MIL 10		-----		
UWG 10		30.1 (15.2-57.5; 1.85)		
TIF 10	Spinosad	0.33 (0.24-0.44; 3.12)		
HAZ 10		0.50 (0.28-0.91; 2.29)		
MIL 10		0.45 (0.26-0.75; 2.07)		
UWG 10		0.45 (0.34-0.60; 2.33)		
TIF 10	Cypermethrin	61.7 (50.1-79.0; 1.99)	36.9 (21.1-58.6; 0.87)	10.9 (8.11-16.4; 1.34)
HAZ 10		60.7 (57.8-71.9; 2.60)	-----	> 20
MIL 10		123.0 (88.2-205.0; 2.74)	-----	14.3 (8.19-54.1; 0.96)
UWG 10		32.2 (27.8-37.6; 2.40)	15.7 (3.14-36.3; 1.07)	5.21 (2.94-9.47; 1.24)

* larval weight: ~35 mg **Adult Vial Test (males and female adults evaluated)

Table 3. Mean susceptibilities of bollworm larvae to MVPII[®], cypermethrin and spinosad (Tracer[®]) following a 96 h exposure period using an insecticide-treated diet bioassay—1990-2010.

Year	LC ₅₀ , ppm (Slope)		
	MVPII [®]	Cypermethrin	Spinosad
1996	38.9 (1.7)	1.40 (2.1)	0.30 (1.6)
1997	68.3 (1.6)	1.31 (2.2)	ND
2003	110 (0.6)*	4.49 (1.8)	0.51 (1.5)
2004	128 (1.1)*	2.63 (3.4)	0.30 (2.1)
2005	122 (0.3)*	1.13 (0.8)	ND
2009	40.1 (0.9)*	8.72 (3.3)	0.54 (3.4)
2010	494 (1.0)*	11.8 (2.6)	0.56 (2.3)

ND = Not Determined

* Data based on tests using neonate larvae

Table 4. Mean susceptibilities of tobacco budworm larvae to MVPII[®], cypermethrin and spinosad (Tracer[®]) following a 96 h exposure period using an insecticide-treated diet bioassay—1995-2010.

Strain	LC ₅₀ , ppm (Slope)		
	MVPII [®]	Cypermethrin	Spinosad
HRV	ND	1.42 (5.2)	0.38 (1.4)
OPS	0.75 (0.7)	5.01 (3.2)	0.14 (3.3)
OPR	ND	5.48 (2.7)	0.37 (2.2)
PYR	1.23 (1.9)	36.5 (2.1)	0.40 (3.4)
1995	0.95 (1.0)	0.46 (1.1)	0.84 (1.7)
1996	9.63 (1.0)	4.32 (3.0)	0.48 (3.1)
1997	8.68 (1.2)	7.55 (2.5)	0.35 (1.8)
1998	ND	12.1 (1.7)	ND
1999	ND	11.5 (0.9)	0.20 (1.9)
2003	1.00 (0.5)*	33.1 (1.4)	0.52 (1.1)
2004	1.20 (1.6)*	33.1 (1.3)	0.40 (1.6)
2005	3.33 (0.5)*	27.6 (1.2)	0.32 (1.2)
2009	2.66 (1.2)*	52.7 (2.2)	0.61 (1.8)
2010	12.2 (1.3)*	69.4 (2.5)	0.43 (2.4)

ND = Not Determined

* Data based on tests using neonate larvae

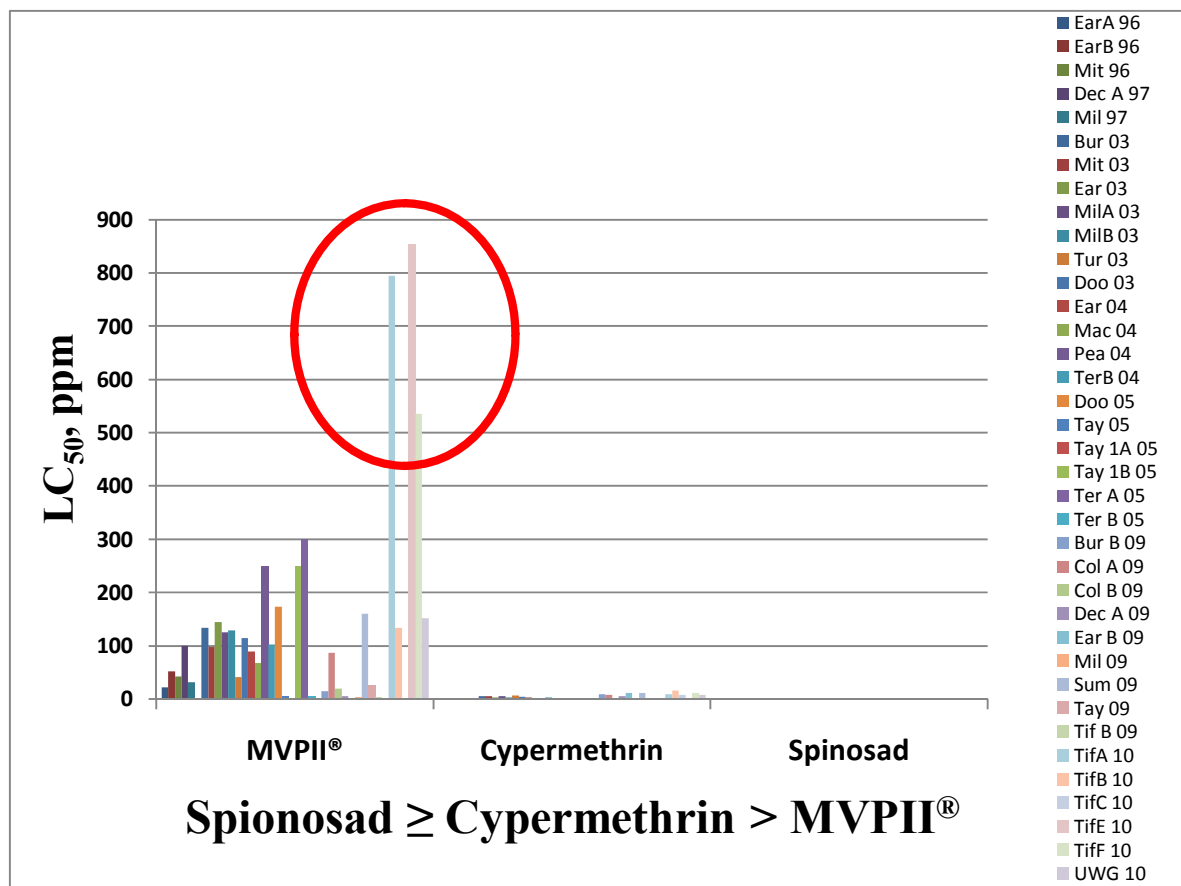


Figure 2. Susceptibilities of field-collected bollworm larvae to MVP II®, spinosad (Tracer®), and cypermethrin using a treated diet bioassay—1996-2010.

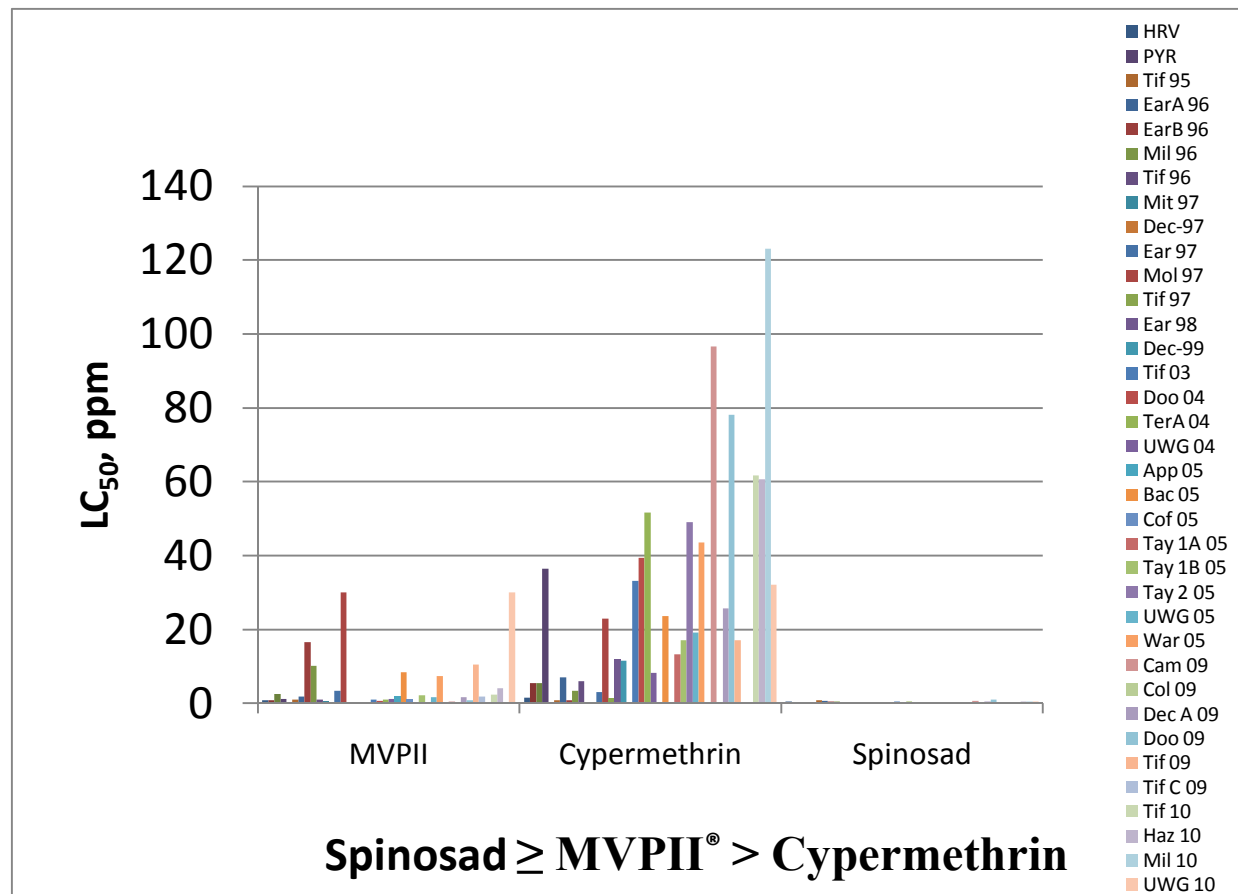


Figure 3. Susceptibilities of field-collected tobacco budworm larvae to MVP II®, spinosad (Tracer®), and cypermethrin using a treated diet bioassay—1996-2010.

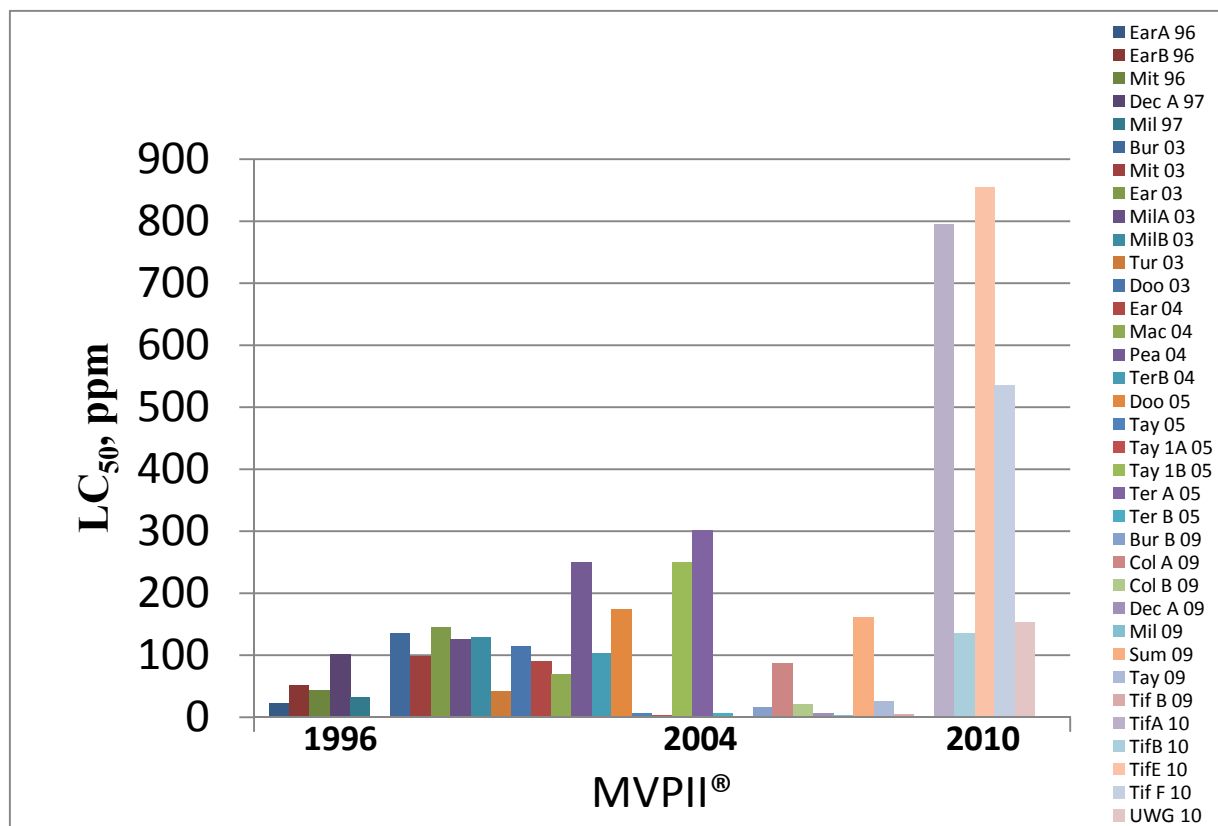


Figure 4. Susceptibilities of field-collected bollworm larvae to MVP II® using a treated diet bioassay—1996-2010.

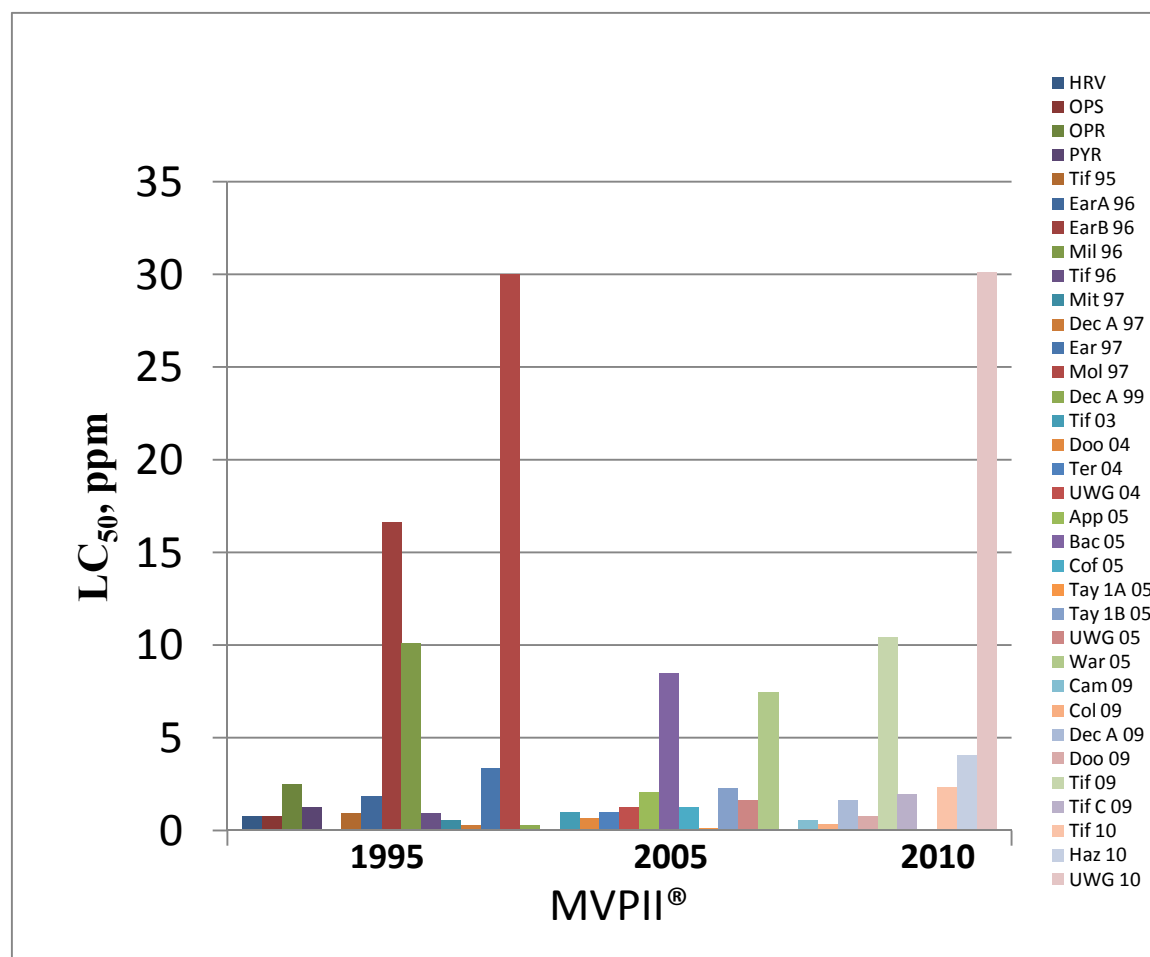


Figure 5. Susceptibilities of field-collected tobacco budworm larvae to MVP II® using a treated diet bioassay—1995-2010.

Although decreases in the susceptibilities of CEW and TBW populations to pyrethroid insecticides were noted throughout the study period and are of considerable concern, data indicated that CEW populations in Georgia remained relatively susceptible to cypermethrin. However, 2010 LC₅₀ values (for both CEW and TBW) were the highest LC₅₀s recorded to date (Tables 1-4; Figures 2, 3, 6, and 7). The average 2009-2010 CEW LC₅₀ value was ca. 7-fold higher than the average 1996-1997 CEW LC₅₀ value and ca. 3-fold higher than the average 2003-2005 CEW LC₅₀ values. The average 2009-2010 TBW LC₅₀ values was ca. 15-fold higher than the average 1995-1996 TBW LC₅₀ value and 2-fold higher than the average 2003-2005 TBW LC₅₀ value. In addition, the average 2009-2010 TBW LC₅₀ value was more than 48-fold higher than the LC₅₀ value obtained for the pyrethroid-susceptible HRV laboratory strain and 1.7-fold higher than the LC₅₀ value obtained for a laboratory-selected, pyrethroid-resistant strain (PYR) (Table 4). As indicated by the previously presented treated diet data, topical application (Tables 1 and 2) and AVT (Tables 1 and 2; Figure 8) data indicated a decrease in the susceptibility of CEW and TBW populations to cypermethrin over time also. Compared to a pyrethroid-susceptible laboratory strain, topical LC₅₀ values for the 2010 CEW populations were 3-fold to 12-fold higher and the percent survival of CEW adults (although comparable to 2009) has steadily risen over the sixteen year period. In field-collected TBW populations, 2010 topical application and AVT cypermethrin data were comparable to data collected during the 2009 season, and an evaluation of LC₉₅ values for cypermethrin against TBW larvae have indicated an annual and sharp increase since the monitoring project began (Figure 9).

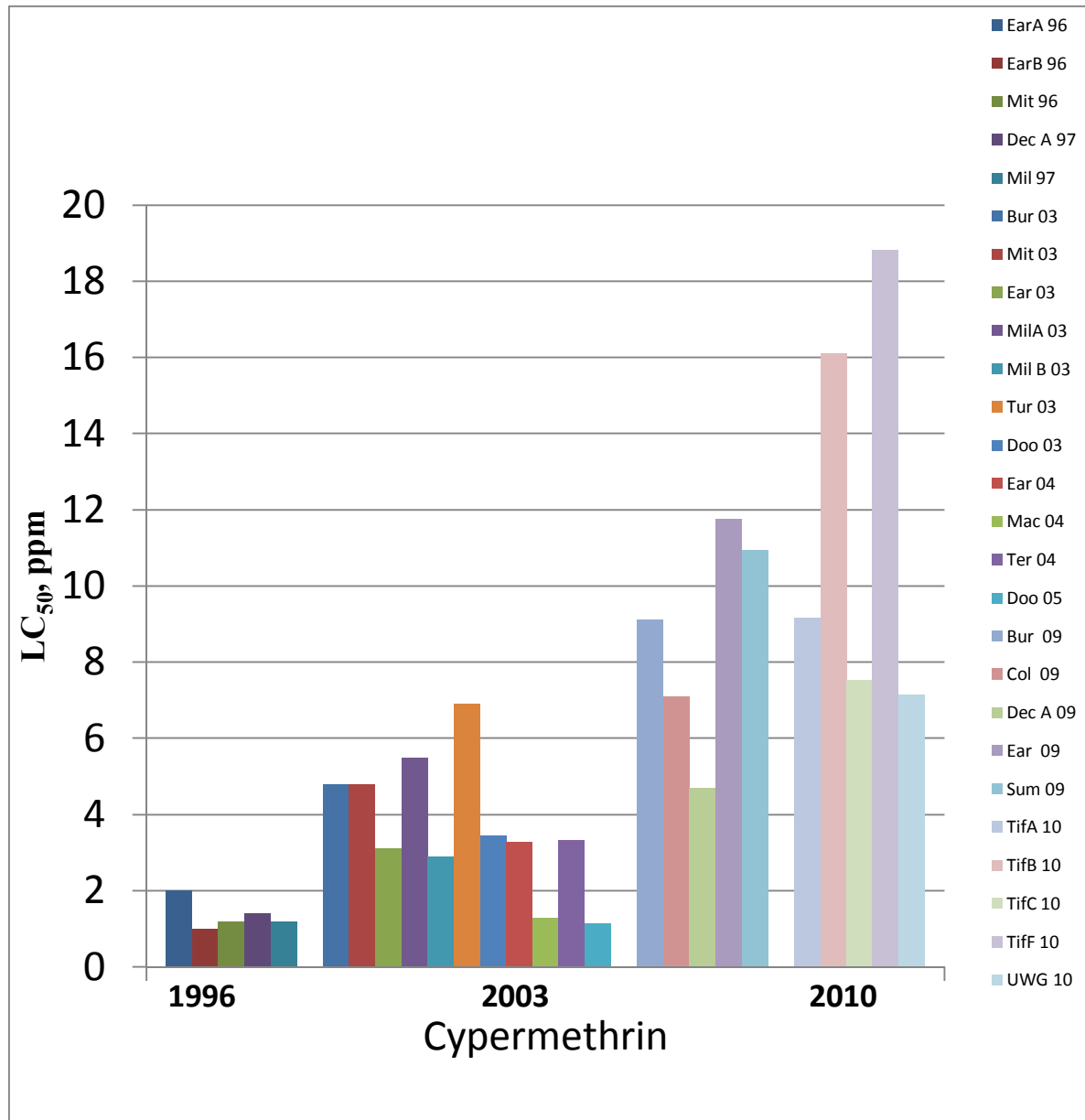


Figure 6. Susceptibilities of field-collected bollworm larvae to cypermethrin using a treated diet bioassay—1996-2010.

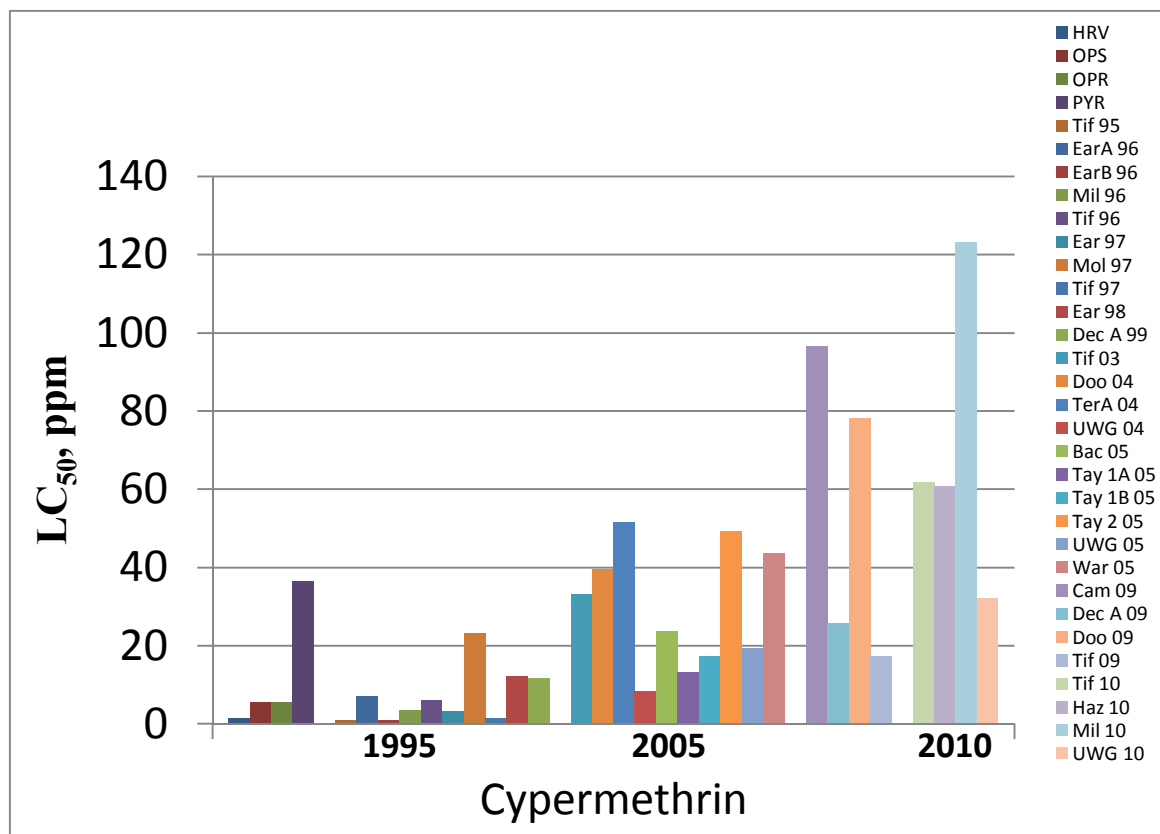


Figure 7. Susceptibilities of field-collected tobacco budworm larvae to cypermethrin using a treated diet bioassay—1995-2010.

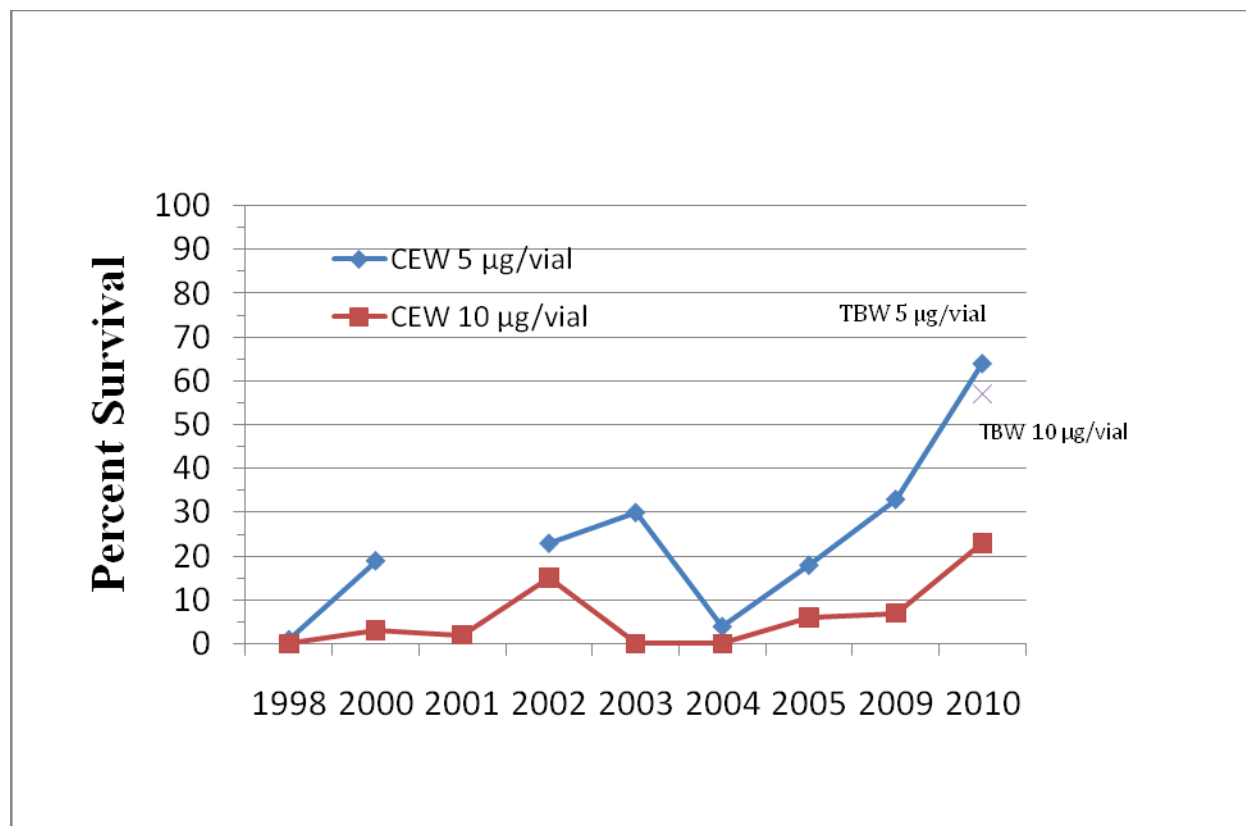


Figure 8. Susceptibilities of bollworm and tobacco budworm adults to cypermethrin using an adult vial test (AVT) bioassay—1998-2009.

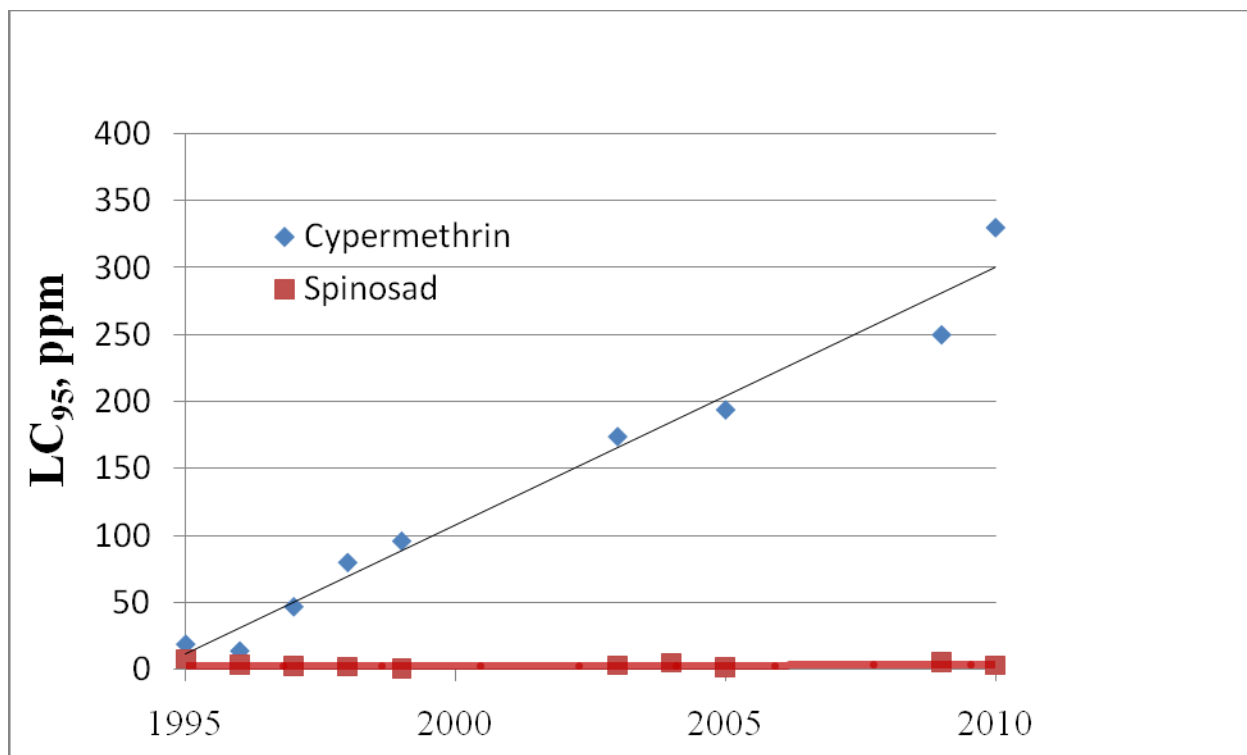


Figure 9. Susceptibilities of bollworm larvae to cypermethrin and spinosad (Tracer[®]) expressed as the LC₉₅ using a treated diet bioassay—1995-2010.

To date, spinosad (Tracer[®]) has remained effective against all strains tested (Tables 1-4; Figures 2, 3, 10, and 11). Mean LC₅₀ values for bollworm larvae (0.56 ppm) and TBW larvae (0.43 ppm) were comparable and have remained stable throughout the sixteen-year study period.

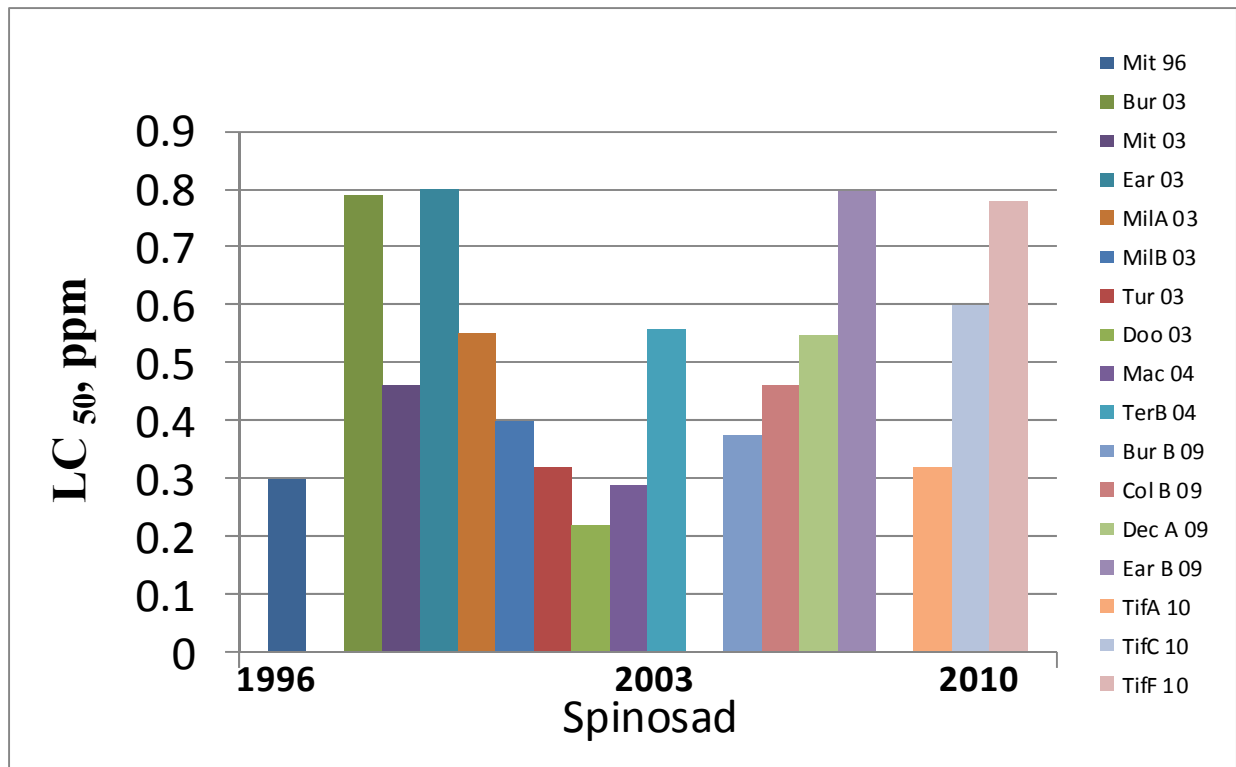


Figure 10. Susceptibilities of field-collected bollworm larvae to spinosad (Tracer[®]) using a treated diet bioassay—1996-2010.

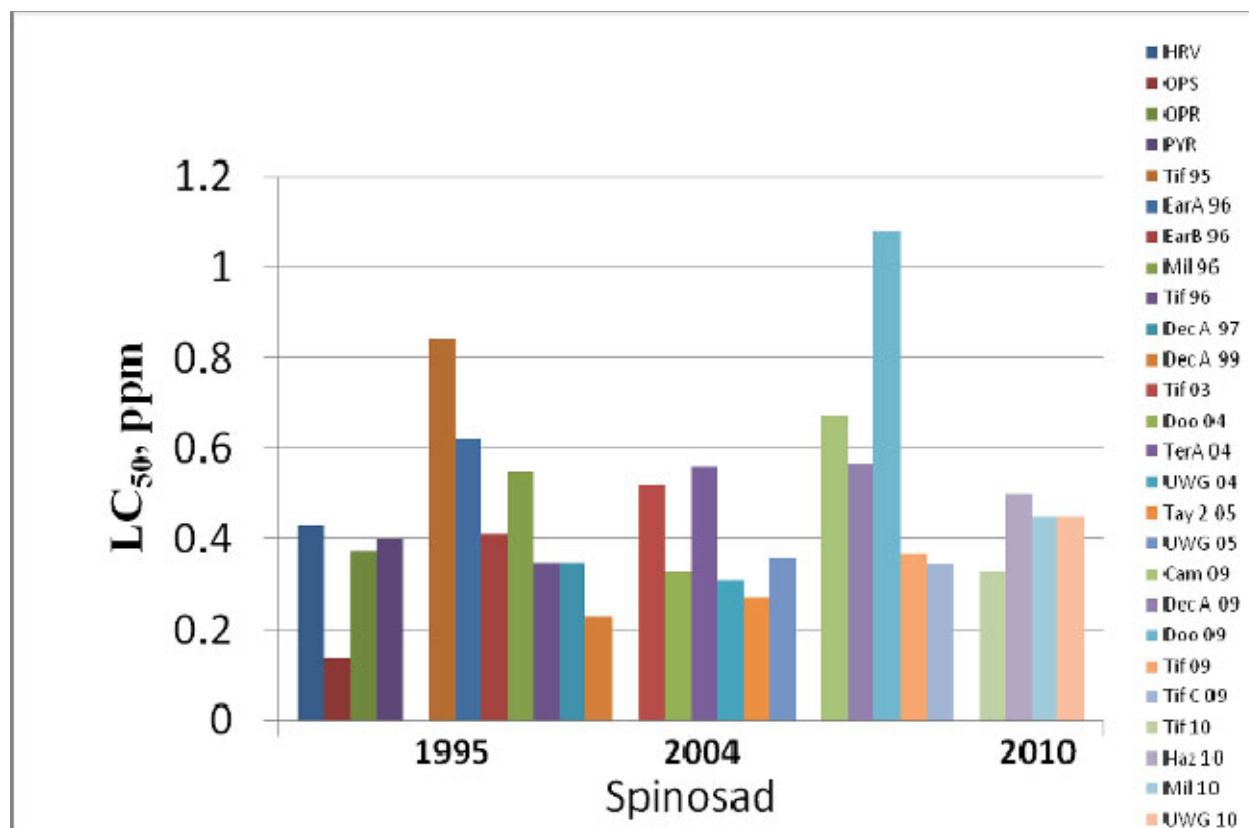


Figure 11. Susceptibilities of field-collected tobacco budworm larvae to spinosad (Tracer®) using a treated diet bioassay—1996-2010.

Summary

Throughout the sixteen year study period bollworm (CEW) and tobacco budworm (TBW) populations in Georgia have remained relatively susceptible to MVP II®. As expected, the data have indicated that CEW larvae were more tolerant to the effects of MVP II® than TBW larvae. CEW and TBW populations have become more resistant to cypermethrin. In 2010, CEW populations were 7 times more resistant to cypermethrin than CEW populations sampled during the mid-1990s and 3 times more resistant to cypermethrin than CEW populations sampled during the mid-2000s. TBW populations collected during the 2010 season were on average 15 times more resistant than TBW populations sampled during the mid-1990s and ca. 2 times more resistant than TBW populations sampled during the mid-2000s. The data indicated that spinosad (Tracer®) has remained effective in the control of CEW and TBW populations in Georgia. There have been no substantial fluctuations in the activity of spinosad against CEW and TBW larvae throughout the study period. In general, the treated diet-96 h activity spectrum for the insecticides tested were as follows: CEW: Spinosad (Tracer®) > Cypermethrin > MVP II®; TBW: Spinosad (Tracer®) > MVP II® > Cypermethrin.

Acknowledgements

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