

VIPCOT™ PERFORMANCE IN THE MID-SOUTH: 2006 – 2007**Scott Martin****Eric Palmer****David Black****Syngenta Crop Protection****Greensboro, NC****David V. Negrotto****David P. Dickerson****Syngenta Biotechnology, Inc.****Research Triangle Park, NC****Abstract**

Syngenta is currently developing VipCot™, a transgenic insect-resistant cotton that expresses dual insect toxins (Vip3A and Cry1Ab). Trials were conducted throughout the cotton belt to test the efficacy of VipCot™ against tobacco budworm, *Heliothis virescens* (Fabricius), and cotton bollworm, *Helicoverpa zea* (Boddie). The efficacy of VipCot™ was compared to that of the conventional variety, Coker 312. VipCot™ provided excellent control of both tobacco budworm and cotton bollworm. These results indicate the high level of insecticidal activity provided by a stack of Vip3A and Cry1Ab toxins. Results from 2006 and 2007 field trials conducted in the Mid-South (LA, MS, AR) will be presented.

Introduction

Currently all available transgenic insect-resistant cotton varieties rely on Cry proteins from *Bacillus thuringiensis* (Bt). Bollgard II® expresses both Cry1Ac and Cry2Ab, whereas Widestrike® expresses Cry1Ac and Cry1F. Thus there is a real need for greater insecticidal protein diversity in the cottonseed marketplace. To meet this need, Syngenta has been developing cotton varieties that express the novel insecticidal protein Vip3A (Estruch et al., 1996). VipCot cotton will comprise Vip3A stacked with the Cry protein Cry1Ab. Vip3A is unrelated to the Cry proteins and targets a distinct binding site in the insect's midgut (Lee et al., 2003; Chen and Lee, 2005). Substantial evidence is available supporting a lack of cross-resistance between Vip3A and Cry proteins (Marcus et al., 2005; Jackson et al., 2006; McCaffery et al., 2006; Fang et al., 2007; Jackson et al., 2007). The combination of Vip3A and Cry1Ab is expected to provide both exceptional lepidopteran insect control spectrum as well as excellent insect resistance management (IRM) attributes.

VipCot field trials testing the agronomic performance and insect efficacy were performed in 2006 and 2007. The field trial program included 35 locations in 2006 and 20 locations in 2007. These trials were run by Syngenta, D&PL & University Cooperators. Efficacy trials were artificially infested with *H. virescens* and *H. zea* where there was a lack on natural lepidopteran insect pressure. Agronomic performance trials were treated as conventional cotton to prevent insect damage and measure plant performance. Larger plot trials (1 – 2 acres) relied on natural infestations (Coker 312 & VipCot only).

Materials and Methods

Several lines were evaluated in the 2006 (Martin et al., 2007) and 2007 field trial programs. All lines were in the Coker 312 background. This report focuses on four of these lines: Coker 312 (non-transgenic), COT67B (Cry1Ab component), COT102 (Vip3A component) and the pyramided combination of COT102 and COT67B (VipCot).

Trials were planted across the US Cotton Belt to evaluate performance in a variety of environmental/growing conditions. This report focuses on trials performed in the Mid-South (Table 1). Results from the South East USA and Texas are presented separately.

Table 1. Trial Locations for 2006 and 2007 Insect Efficacy Trials – Mid-South

Bosier City, LA (M)	2006
Bosier City, LA (M)	2007
Winnsboro, LA (L)	2006
Winnsboro, LA (L)	2007
Leland, MS	2006
Stoneville, MS	2007
Scott, AR	2006
Scott, AR	2007

Entomological Assessments:

Natural infestations of *H. virescens* / *H. zea*: All entries were assessed at ~ 7 day intervals during the entire period when there was significant insect pressure on the trial. At each evaluation, 100 randomly selected squares and bolls were assessed per plot. Each structure was rated for feeding damage. A damage rating was based on fruiting body penetration, rather than on superficial feeding damage. The data provided were the number of damaged structures per 100 structures.

Artificial infestation of *H. virescens* / *H. zea*: Artificial infestations were only carried out if less than 10% square damage due to the relevant species was observed in Coker 312 by ~ 3 weeks after pinhead square. Ratings were carried out 3 and 7 days after the artificial infestation. At each evaluation, 100 randomly selected squares and bolls were assessed per plot. Each structure was rated for feeding damage. A damage rating was based on fruiting body penetration, rather than on superficial feeding damage. The data provided were the number of damaged structures per 100 structures.

Results

Agronomic results obtained in trials from 2006 were reported (Mahaffey 2007). Analyses of the 2007 Agronomic trials are not complete.

In the tobacco budworm (TBW) plots (Figure 1), Coker 312 showed a high level of square damage. Overall, VipCot showed less square damage than both parent lines, COT102 (Vip3A) and COT67B (Cry1Ab). The missing locations either had no natural TBW pressure, the artificial infestations did not succeed or the data were not available.

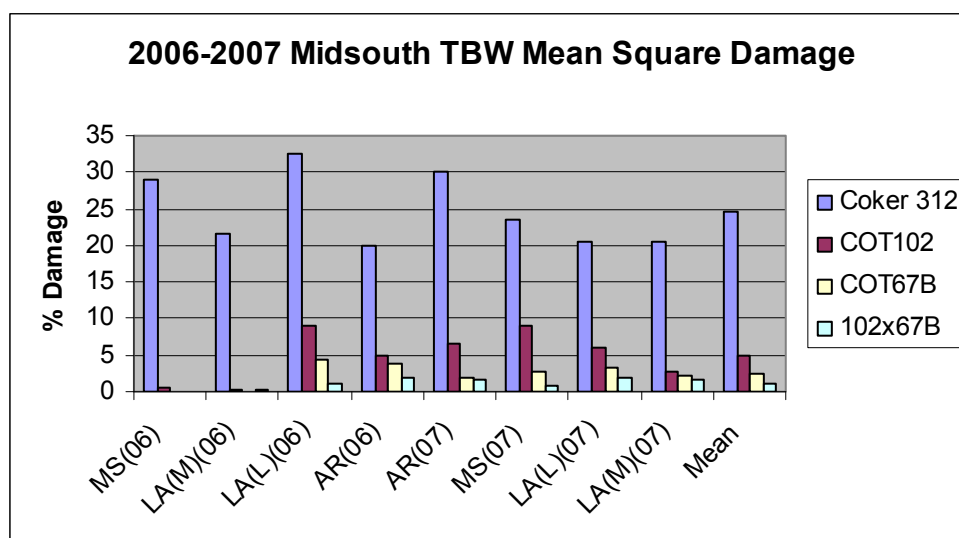


Figure 1.

In the cotton bollworm (CBW) plots (Figure 2), Coker 312 showed a high level of square damage. Overall, VipCot showed less square damage than both parent lines, COT102 (Vip3A) and COT67B (Cry1Ab). The missing locations either had no natural CBW pressure, the artificial infestations did not succeed or the data were not available.

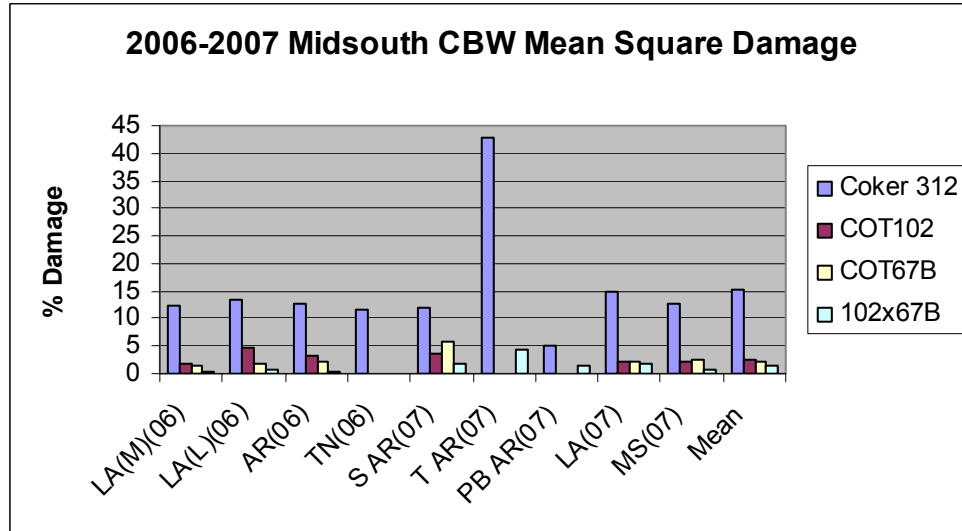


Figure 2.

In the cotton bollworm (TBW) plots (Figure 3), Coker 312 showed a high level of boll damage. Overall, VipCot showed less square damage than both parent lines, COT102 (Vip3A) and COT67B (Cry1Ab). The missing locations either had no natural TBW pressure, the artificial infestations did not succeed or the data were not available.

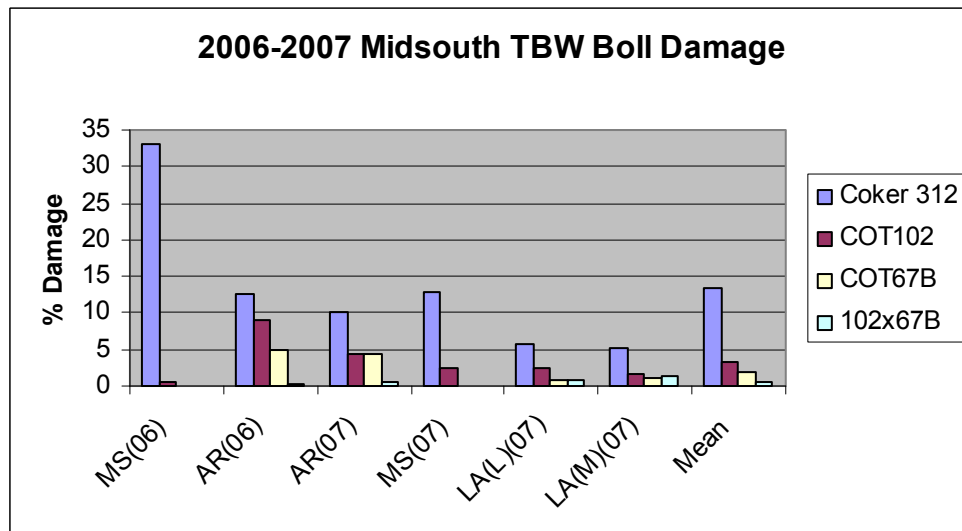


Figure 3.

In the cotton bollworm (CBW) plots (Figure 4), Coker 312 showed a high level of boll damage. Overall, VipCot showed less boll damage than both parent lines, COT102 (Vip3A) and COT67B (Cry1Ab). The missing locations either had no natural CBW pressure, the artificial infestations did not succeed or the data were not available.

Figure 4.

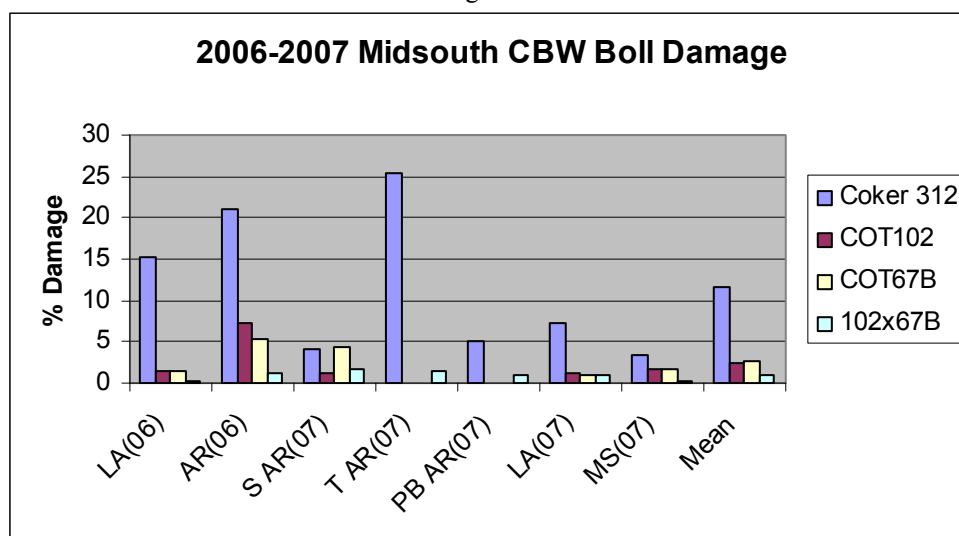


Figure 4.

Conclusions

VipCot provided excellent control of both TBW (*H. virescens*) and CBW (*H. zea*), resulting in little or no square or boll damage. Additionally, VipCot provided levels of control better than either of the parental lines included in these trials. These data confirm VipCot cotton has excellent activity against these key heliothine pests in the USA.

The risk of cross-resistance developing between Vip3A and Cry toxins is extremely low. Vip3A represents an excellent partner for Cry toxins in stacked cotton product offerings due to having exceptional efficacy, no adverse agronomic effects, and excellent insect resistance management properties

Literature Cited

- Chen, E. and M. Lee. 2005. *In vitro* cross-resistance studies with Vip3A to support the IRM strategy for VipCot™. In Proc. 2005 Beltwide Cotton Conf. National Cotton Council, Memphis, TN.
- Estruch, J. J., G. W. Warren, M. A. Mullins, G. J. Nye, J. A. Craig, and M. G. Koziel. 1996. Vip3A, a novel *Bacillus thuringiensis* vegetative insecticidal protein with a wide spectrum of activities against lepidopteran insects. Proceedings National Academy of Science USA. 93: 5389-5394.
- Fang, J., X. Xu, P. Wang, J. Zhao, A. M. Shelton, J. Cheng, M.-G. Cheng and Z. Shen. 2007 Characterization of chimeric *Bacillus thuringiensis* Vip3 toxins. Applied and Environmental Microbiology, February 2007, p. 956-961, Vol. 73, No. 3.
- Jackson, R.E., M.A. Marcus, F. Gould, J.R. Bradley, A.R. McCaffery and J. Van Duyn. 2006. Cross-resistance levels between the *Bacillus thuringiensis* proteins Cry1Ac and Vip3A in *Heliothis virescens*. In Proceedings, 2006 Beltwide Cotton Conference. National Cotton Council, Memphis, TN.
- Jackson, R. E., M. A. Marcus, F. Gould, J. R. Bradley and J. Van Duyn. 2007. Cross-resistance responses of Cry1Ac-selected *Heliothis virescens* (Lepidoptera: Noctuidae) to the *Bacillus thuringiensis* protein Vip3A. Journal of Economic Entomology. Volume 100, Number 1, February 2007, pp. 180-186(7).

Lee, M.K., F.S. Walters, H. Hart, N. Palekar, and J.S. Chen. 2003. The mode of action of *Bacillus thuringiensis* vegetative insecticidal protein Vip3A differ from that of Cry 1Ab delta-endotoxin. *Appl. and Environ. Microbiol.* 69: 4648-4657.

Mahaffey, J., J. Burgess, K. Howard, and T.A. Kerby. Agronomic Evaluations of VipCot™ Containing Cotton Genotypes: Vip3A, Cry1Ab, and Stacked Combinations. *In Proc. 2007 Beltwide Cotton Conf. National Cotton Council, Memphis, TN.*

Marcus, M., J.R. Bradley, F.L. Gould, and J.W. Van Duyn. 2005. Cross-resistance evaluations of Cry1Ac tolerant *Heliothis virescens* strains to the novel insecticidal protein Vip3A, *In Proceedings, 2005 Beltwide Cotton Conference. National Cotton Council, Memphis, TN.*

Martin, S., B. Minton, J. Hamill, J. Holloway, D. Black, D. Porterfield and D. Dickerson. 2007. Field evaluation of VipCot™ cotton against Heliothines under natural and artificial infestations, *In Proceedings, 2007 Beltwide Cotton Conference. National Cotton Council, Memphis, TN.*

McCaffery, A., M. Capiro, R. Jackson, M. Marcus, T. Martin, D. Dickerson, D. Negrotto, D. O'Reilly, E. Chen, and M. Lee. 2006. Effective IRM with a novel insecticidal protein, Vip3A, p. 1229-1235. *In Proc. 2006 Beltwide Cotton Conf. National Cotton Council, Memphis, TN.*